

Ultra-weak photon emission: a nondestructive detection tool for food quality and safety assessment

Mohammad Amin Nematollahi^{1,*}, Zahra Alinasab², Seyed Mehdi Nassiri¹, Amin Mousavi Khaneghah^{3,*}

¹Department of Biosystems Engineering, College of Agriculture, Shiraz University, Shiraz, Iran; ²Department of Medical Physics, Isfahan University of Medical Sciences, Isfahan, Iran; ³Department of Food Science, Faculty of Food Engineering, University of Campinas (UNICAMP), São Paulo, Brazil

***Corresponding Authors:** Mohammad Amin Nematollahi, Department of Biosystems Engineering, College of Agriculture, Shiraz University, Shiraz, Iran. Email: manema@shirazu.ac.ir; Amin Mousavi Khaneghah, Department of Food Science, Faculty of Food Engineering, University of Campinas (UNICAMP), Rua Monteiro Lobato, 80. Caixa Postal: 6121.CEP: 13083-862, Campinas, São Paulo, Brazil. Email: mousavi@unicamp.br

Received: 11 June 2020; Accepted: 26 September 2020; Published: 16 October 2020

© 2020 Codon Publications



REVIEW ARTICLE

Abstract

A new aspect covering interactions between cells and their surroundings via electromagnetic waves was introduced by applying ultra-weak photon emission (UPE). The UPE originates from the relaxation of electronically excited species resulting from oxidative metabolic processes and oxidative stress associated with reactive oxygen species (ROS). The ROS plays a critical role in the quality of foods, and their determination is of extreme importance. The ROS and the intensity of the UPE have significantly correlated. The UPE can be effectively monitored by specific instruments such as photomultiplier tube and charged-coupled devices. The current review is devoted to providing an overview of the quality of food products by the aid of UPE via evaluating the correlations between UPE and food quality indices. In this regard, the UPE can be utilized in food quality as a real-time, noninvasive, and nondestructive technique without complex instruments. However, the implementation of the UPE method for evaluation of food quality needs further investigations.

Keywords: defense mechanisms; food quality assessment; oxidative stress; reactive oxygen species; ultra-weak photon emission

Introduction

All known biological systems possess an active oxidative metabolism or stress that involves oxidation reactions in which reactive oxygen species (ROS) play a critical role. These ROS can efficiently react with biomolecules in organisms, resulting in the synthesis of unstable intermediates. The decomposition of these intermediates mostly leads to the formation of unstable excited electron species. During this formation, a tiny amount of light is generated, and monitoring of this light can give essential information about the organism's oxidative state (Cifra and Pospíšil, 2014; Pospíšil *et al.*, 2014).

Investigations regarding cellular communication through ultra-weak photon emission (UPE) started in the 1910s. At that time, a scientist named Alexander Gurwitsch, after conducting various experiments revealed that two separate series of onion root cell cultures, adjacent to each other, could pose some influences on each other regarding cell division and multiplication rate (Bischof, 2003; Prasad *et al.*, 2014; Scholkmann *et al.*, 2013).

The emission of light by various living organisms was demonstrated by several previous investigations (Burgos *et al.*, 2017; Cifra and Pospíšil, 2014; de Mello Gallep and Robert 2020; Esmaeilpour *et al.*, 2020; Jia *et al.*, 2020;

Prasad *et al.*, 2014; Prasad *et al.*, 2020; Van Wijk *et al.*, 2001). According to literature, the concept of UPE was introduced in a variety of terms such as “biophotons,” “ultra-weak emission,” “self-bioluminescent emission,” “photoluminescence,” “delayed luminescence,” “ultra-weak luminescence,” “spontaneous chemiluminescence,” “endogenous bioluminescence,” and “biochemiluminescence” (Salari *et al.*, 2011; Shanei *et al.*, 2017).

As very active and unstable compounds, free radicals are referred to as atoms, molecules, or ions with unpaired electrons (Mayorga Burrezo *et al.*, 2019). In this regard, the oxygen radicals are classified as free radicals which can be produced continuously in all living organs with destructive effects on cellular proteins, lipids, and, most notably, DNA that may lead to carcinogenesis (Saikolappan *et al.*, 2019). The ROS can be classified into two groups as radical and non-radical species (Gill and Tuteja, 2010; Pospíšil *et al.*, 2019). The ROS can react with biomolecules, such as lipids, nucleic acids, and proteins, to cause a deformity and finally increase their levels of energy. Consequently, this reaction creates electron excitation, with later electron's transition from a singlet-triplet state to the base state with photon emission, usually called UPE (Pospíšil *et al.*, 2014).

The formation of ROS in the food and agricultural industry must be monitored as it is strongly related to public health and may cause an economic burden at a global level. ROS production can be associated with monitoring plant response to pathogens, drought stress, flooding stress, salt stress, and herbicides among agriculture products. Currently, the evaluation of UPE as a robust, real-time, inexpensive, nondestructive, and noninvasive tool to monitor oxidative reactions among several scientific fields, such as medical, pharmaceutical, biological, environmental, agricultural, and food products, is the point of interest. A probable correlation between UPE and food quality indices can be proposed (Gałazka-Czarnecka *et al.*, 2019; Sun *et al.*, 2019). Therefore, the UPE as a diagnostic tool to monitor agriculture processes can be considered for further developments (Cifra and Pospíšil, 2014; Guo *et al.*, 2017; Inagaki *et al.*, 2008; Moraes *et al.*, 2012; Prasad and Pospíšil, 2011).

Due to the rapid growth of the world population, food security, safety, and quality are important issues that should be considered severe challenges (Cheeseman, 2016; Godfray *et al.*, 2010; McCarthy *et al.*, 2018; Prosekov and Ivanova, 2018). Destructive methods are widely employed to evaluate food quality, but they are usually more labor-intensive and time-consuming, which may harm the material. In contrast, the non-destructive methods allow the measurement of different food quality attributes without affecting physical

structure and quality. Therefore, the use of nondestructive methods has attracted many researchers (El-Mesery *et al.*, 2019; Magwaza *et al.*, 2013). Traditional non-destructive techniques, such as machine vision, hyper-spectral imaging, near-infrared (IR) spectroscopy, electronic nose, electronic eye, electronic tongue, ultrasound measurements, and acoustic emission measurements, have been employed to assess the quality of food and agricultural products (El-Mesery *et al.*, 2019; Giovenzana *et al.*, 2017; Kheiralipour *et al.*, 2016; Omar and MatJafri, 2013; Schinabeck *et al.*, 2018; Zhong and Wang, 2019).

Currently, the application of UPE in food quality is a hot topic, and investigations are still ongoing regarding measuring food quality indices. However, to the best of our knowledge, no overview of this subject in the food quality area has been provided. Therefore, this article was undertaken to provide an overview considering the measurement of ROS production by UPE in food quality assessment. In this context, the definition, sources of generation, detection mechanisms, and applications of UPE in agricultural products are pinpointed.

Ultra-Weak Photon Emission

UPE definition

In addition to chemical signal transduction pathways, the communication between living beings can be carried out through electromagnetic waves (Van Wijk, 2001). In this regard, they could emit light either spontaneously or coherently, which is different from fluorescence, phosphorescence, and conventional bioluminescence (Cifra and Pospíšil, 2014; Shanei *et al.*, 2017). The spontaneous emission can occur without an external excitation or any pre-illumination. The living organisms have a nonexponential decay of UPE after exposure to external light (Rafii-Tabar and Rafieiolhosseini, 2015). The coherent emission is another aspect of UPE, defined as a state of light in which waves can interfere constructively and form interference patterns (Gu, 1999).

As mentioned earlier, the oxidation of biomolecules during cellular metabolism leads to UPE. It was also reported that an organism's DNA could act as a source of UPE (Prasad *et al.*, 2014).

Induced UPE can be originated from various oxidative factors, mainly biotic and abiotic stresses. The biotic factors include bacterial (Mansfield, 2005), viral (Kobayashi *et al.*, 2006), fungal (Rastogi and Pospíšil, 2012), and herbivorous stress (Yoshinaga *et al.*, 2006). The abiotic stresses arise from factors such as the surrounding environment (Münzel *et al.*, 2018), mechanical damage

(Liang *et al.*, 2019), undesired temperature (Ahammed *et al.*, 2019), light (Nakashima *et al.*, 2017), and ionizing radiation (Singh *et al.*, 2017). All these factors increase oxidative damages because of excessive production of ROS. The UPE possesses a spectral range varying from 200 to 800 nm with fragile intensity (few to hundreds of photons per cm²) (Yang *et al.*, 2017). It is interesting to note that such intensity of radiation is equivalent to look at candlelight with naked eyes from a distance of nearly 24 km (Bischof, 2005), which elucidates the difficulty of capturing these signals.

UPE detection

The photographic containers and tubes with a particular sensibility to ultraviolet (UV) rays were the first employed devices for UPE detection, capturing waves in the UV range (Cifra *et al.*, 2011). However, the intensity of these waves was very weak for such detection with available detectors. Therefore, their detection was postponed for many years. After some advancements in technology, several suitable devices were introduced to detect UPE, for example, avalanche photodiodes (APD), photodiode arrays (PDA), charged-coupled devices (CCD), microchannel plate (MCP), visible light photon counters (VLPC), superconducting tunnel junctions (STJ), hybrid photon detector (HPD), photo multiplier-tube (PMT), and channel photomultiplier (CPM). PDAs, CCDs, and MCPs, are used for spatial (2-dimensional [2D]) resolution. Considering the devices introduced for 1D resolution, PMTs deal with a characteristic photon density ranging from a few to up to some hundred photons per square centimeter per second. Therefore, they remain a suitable choice for UPE detection. After PMT development in the 1950s, detection of this light achieved notable improvement, and measurements became accurate (Bischof, 2005; Rahnama *et al.*, 2011). At this stage, scientists discovered that UPE was also emitted in the

visible range and UV range. Up to now, PMT and CCD cameras have been used widely for the detection of UPE (Cifra *et al.*, 2011). While the former enhances light radiation up to 100 times, the latter detects this light in two dimensions as a highly sensitive photon detector. The MCP devices are sensitive compared with PDA and CCD (Madl, 2014; Ortega-Ojeda *et al.*, 2018).

Effect of ROS on Plants

Oxidative stress

Oxidative stress is defined as “an imbalance between oxidant production by free radicals and the antioxidant capacity of the cell” (Sies *et al.*, 2017), which causes severe adverse effects on the growth and productivity of plants. The biotic, abiotic, and stress conditions (Pitzschke *et al.*, 2006) are demonstrated in Figure 1. ROS are free radicals and play a critical role in oxidative stress in which their accumulation in the plant cell leads to damage to some organelles. Besides, along with ROS, reactive nitrogen and sulfur species play an essential role in the cell's oxidative stress development.

ROS formation

Reactive oxygen species in all aerobic organisms, as well as plants, are continuously formed as a toxic by-product as a result of aerobic metabolism, while they also can be originated from various enzymatic and nonenzymatic processes as well as two biotic and abiotic factors (Bailey-Serres and Mittler, 2006; Gupta *et al.*, 2015). While the sources of ROS in plant cells are located in chloroplasts, mitochondria, peroxisomes, endoplasmic reticulum, apoplast, plasma membranes, and cell wall (Abouzari and Fakheri, 2015), some ROS can be detoxified by some enzymatic and nonenzymatic mechanisms

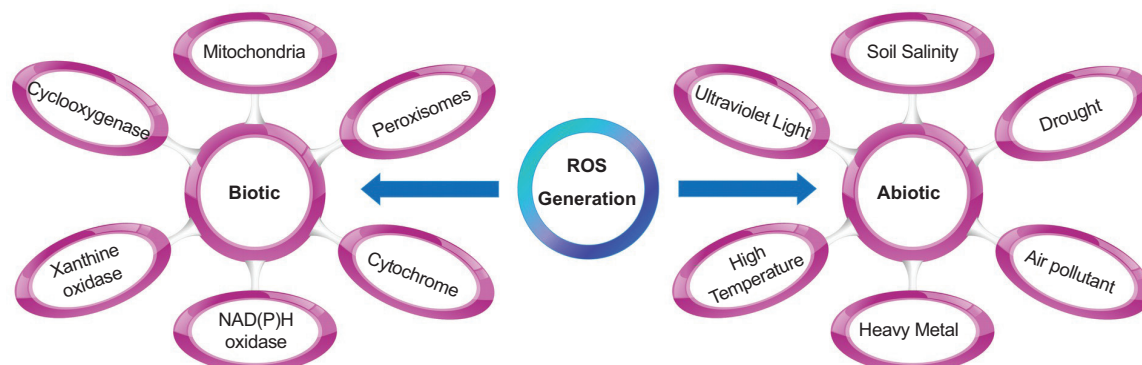
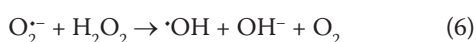
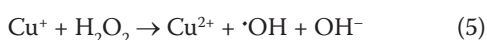
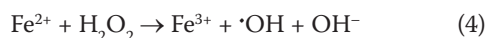
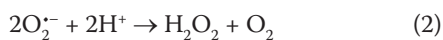


Figure 1. Sources of ROS generation. ROS, reactive oxygen species.

(Ahmad, 2013). Figure 2 demonstrates the radical and non-radical forms of ROS.

The reactions for significant ROS generation can be summarized as follows:



The one-electron reduction of molecular oxygen is responsible for forming high-reactive superoxide radicals (Equation 1). This reduction occurs in mitochondria, chloroplasts, and peroxisomes (Pospíšil *et al.*, 2019). Unlike the superoxide radical, hydrogen peroxide, which is formed through a dismutation reaction by superoxide dismutase (SOD) enzyme, is relatively stable and less reactive (Equation 2) (Battin and Brumaghim, 2009). Hydrogen peroxide can also be produced by different enzymes, such as glycolate oxidase, L-amino acid oxidase, and urate oxidase (Battin and Brumaghim, 2009; Thannickal and Fanburg, 2000). Singlet oxygen, a non-radical, is an excited state of O_2 , which is not very reactive in its ground state (Equation 3). The chlorophyll and their precursors performed singlet oxygen's primary production (Krieger-Liszka, 2005; Tripathy and Oelmüller, 2012). In the presence of transition metals (iron $[\text{Fe}^{2+}]$ or copper $[\text{Cu}^{2+}]$ ions), the highly reactive hydroxyl radical is formed (Equations 4 and 5) through the Fenton reaction (Battin and Brumaghim, 2009; Janků

et al., 2019). Another source of hydroxyl radical is the Haber–Weiss reaction (Equation 6). The Haber–Weiss cycle is a two-step reaction. In the first step, the ferric (Fe^{3+}) ion reduction into the ferrous (Fe^{2+}) ion occurs via reaction with superoxide radical. The second step is the Fenton reaction (Equations 4 and 5). The first and second steps' net reaction is the Haber–Weiss reaction (Kehrer, 2000).

Under normal conditions, the ROS production rate in cells is low (240 $\mu\text{mol/s}$ and 0.5 $\mu\text{mol H}_2\text{O}_2$ at a steady-state level), and the ROS generation is generally in balance with antioxidant capacity. When the oxidative stress exceeds the available antioxidants, the ROS generation's rate increases (240–720 $\mu\text{mol/s}$ and 5–15 $\mu\text{mol H}_2\text{O}_2$ in a steady-state level), which consequently, due to further accumulation, causes cell death when some adverse environmental factors perturb the balance between the rate of production and scavenging of ROS, the intracellular levels of ROS may rapidly rise (Pitzschke *et al.*, 2006; Tsugane *et al.*, 1999). Some defense mechanisms involved antioxidant agents that work hand in hand to reduce undesirable phenomenon (Racchi, 2013).

Antioxidant defense mechanisms

An antioxidant is a substance in low concentration that significantly inhibits oxidation or delays oxidation with different mechanisms (Mousavi Khaneghah, 2016). Among them, oxygen removal or localized oxygen reduction, removal of metal catalytic Cu^{2+} and Fe^{2+} , removal of ROS such as O_2 , H_2O_2 , and chain reaction interruptions, increase in the rate of ROS scavenging, acceleration of recovery of damaged cell structures, and enhancement of absorbed energy heat dissipation are mentioned in literature. Generally, an antioxidant's capacity to neutralize ROS action and free radicals depends on various factors

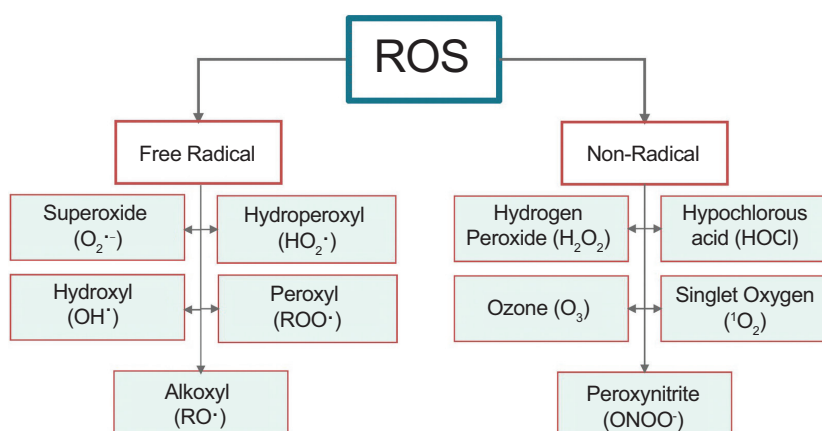


Figure 2. Radical and non-radical forms of ROS. ROS, reactive oxygen species

such as activity, interaction with other antioxidants (synergistic), absorption, distribution, and metabolism of antioxidants (Pitzschke *et al.*, 2006). Antioxidant systems can be classified into enzymatic and nonenzymatic groups (Table 1) (Caverzan *et al.*, 2019; Karuppanapandian *et al.*, 2011).

Recently, several studies have been carried out to find the effects of various antioxidants on UPE. The experimental evidence showed that the antioxidants suppressed UPE in various living organisms. For instance, the UPE from human skin was suppressed by three antioxidants (d- δ -tocopherol sodium, L-glutathione, and L-ascorbate) (Tsuchida *et al.*, 2019). Different antioxidants (α -tocopherol, glutathione, +6 ascorbate, and coenzyme Q10) notably decreased the UPE from the human skin (Rastogi and Pospíšil, 2011). In another research, the results showed that the topical application of oligomeric proanthocyanidins (antioxidants) significantly reduced the UPE from the human skin (Van Wijk *et al.*, 2010). Similar research on humans (Egawa *et al.*, 1999; Sauermann *et al.*, 1999) and mouse skins (Evelson *et al.*, 1997) were conducted to find the effect of antioxidants UPE. It has also been shown that the UPE from radish root cells was considerably suppressed by different amounts of sodium ascorbate and cysteine (Rastogi and Pospíšil, 2010). The ROS induced in rice cells by *N*-acetylchitoooligosaccharide, and consequently UPE, was also highly suppressed by the addition of diphenyl iodonium as a ROS scavenger (Kageyama *et al.*, 2006).

Correlation between ROS and UPE

All living systems are connected to their surroundings through the UPE exchange, while the spontaneous UPE is originated from the transition of electronically excited species to the ground state formed during oxidative metabolic processes (Pospíšil *et al.*, 2014). Cyclo-addition of $^1\text{O}_2$ or the hydrogen abstraction $\cdot\text{HO}$ is two mechanisms for oxidation of biomolecules among oxidative metabolic processes. The hydrogen abstraction from proteins, nucleic acids, and lipids by $\cdot\text{HO}$ can result in an alkyl radical ($\text{R}\cdot$), which could react with O_2 and produce peroxy radical ($\text{ROO}\cdot$). The cyclization of $\text{ROO}\cdot$ and the recombination of two $\text{ROO}\cdot$ lead to high-energy intermediates dioxetane (ROOR) and tetroxide (ROOOOR), respectively. The production of dioxetane is also performed by cycloaddition of singlet oxygen. The electronic species, such as triplet-excited carbonyls ($^3\text{R}=\text{O}^*$), singlet ($^1\text{P}^*$), and triplet pigment ($^3\text{P}^*$), and $^1\text{O}_2$ are formed due to the decomposition of tetroxide and dioxetane (Fedorova *et al.*, 2007; Pospíšil *et al.*, 2014; Yang *et al.*, 2015).

The spectrum of the spectrum associated with photon emission of $^3\text{R}=\text{O}^*$ (350–550 nm) is near UV and

Table 1. Classification of important enzymatic and nonenzymatic antioxidants.

ROS scavenging by antioxidant enzymes	
Enzymes	Reactions
SOD	$2\text{O}_2^- + 2\text{H}^+ \rightarrow \text{H}_2\text{O}_2 + \text{O}_2$
CAT	$2\text{H}_2\text{O}_2 \rightarrow 2\text{H}_2\text{O} + \text{O}_2$
POD	$\text{H}_2\text{O}_2 + \text{GSH} \rightarrow \text{H}_2\text{O} + \text{GSSG}$
APX	$\text{H}_2\text{O}_2 + \text{ASCA} \rightarrow 2\text{H}_2\text{O} + \text{DHA}$
MDHAR	$2\text{MDHA} + \text{NAD(P)H} \rightarrow 2\text{ASCA} + \text{NAD(P)}$
DHAR	$\text{DHA} + 2\text{GSH} \rightarrow \text{ASCA} + \text{GSSG}$
GR	$\text{GSSG} + \text{NAD(P)H} \rightarrow 2\text{GSH} + \text{NAD(P)}^+$
GPX	$\text{H}_2\text{O}_2 + 2\text{GSH} \rightarrow 2\text{H}_2\text{O} + \text{GSSG}$
ROS scavenging by antioxidant nonenzymes	
Nonenzymes	Reactions
AscA	Detoxifies H_2O_2 , O_2^- and $\cdot\text{OH}$
GSH	Substrate for various PODs, GSTs, and GR. Detoxifies H_2O_2 , O_2^- , and $\cdot\text{OH}$
TOCs	Protects membrane lipids from peroxidation, detoxifies lipid peroxides, and quenching $^1\text{O}_2$
CARs	Quench $^1\text{O}_2$
Flavonoids	Can directly scavenge H_2O_2 and $\text{OH}\cdot$

AscA: ascorbic acid; APX: ascorbate peroxidase; CARs: carotenoids; CAT: catalase; DHA: dehydroascorbate; DHAR: dehydroascorbate reductase; GPX: guaiacol peroxidase; GR: glutathione reductase; GSH: glutathione; GSSG: oxidized glutathione; GSTs: glutathione-S-transferases; MDHA: monodehydroascorbate; MDHAR: monodehydroascorbate reductase; PODs: peroxidases; SOD: superoxide dismutase; TOCs: tocopherols.

blue-green regions of visible light (Fedorova *et al.*, 2007). The spectrum range of singlet- and triplet-excited pigments belongs to green-red (550–750 nm) and red–near IR (750–1000 nm), respectively (Pospíšil *et al.*, 2014; Sauermann *et al.*, 1999). The photon emissions of monomol $^1\text{O}_2$ and dimol are close to IR (at 1270 nm) and the visible light (at 634 and 703 nm), respectively (Adam *et al.*, 2005; Pospíšil *et al.*, 2014).

UPE Application in Food and Agriculture

As already discussed, UPE is produced and released by ROS and received by biological systems, especially in agriculture. However, how UPE is emitted and received in intercellular and intracellular interactions is still a significant issue. Since the detection and analysis of the UPE spectrum are simple, available, inexpensive, and noninvasive, it can be used in different fields.

As mentioned earlier, most experiments demonstrate the impact of UPE in the field of agriculture, which can be used for the detection of pathogens in plants (Bennett

et al., 2005; Iyozumi *et al.*, 2002; Kageyama *et al.*, 2006; Kobayashi *et al.*, 2006; Makino *et al.*, 1996; Mansfield, 2005; Montillet *et al.*, 2005; Rastogi and Pospíšil, 2012), drought stress (Guo and Tan, 2013; Kausar *et al.*, 2012; Komatsu *et al.*, 2014; Ohya *et al.*, 2000), salinity (Ohya *et al.*, 2000), flooding stress (Kamal and Komatsu, 2016; Kausar *et al.*, 2012; Khatoon *et al.*, 2012; Komatsu *et al.*, 2014), and herbicides (Inagaki *et al.*, 2007, 2008, 2009; Kato *et al.*, 2014; Nukui *et al.*, 2013).

Food quality and safety with UPE

As stated above, the studies on UPE in the areas of food quality are limited. The aim is to review the useful, relevant documents to find the relation between food quality and safety with UPE (Table 2).

For better clarity, the extensive description of each study (Table 2) follows.

The UPE method was employed to assess the quality of milk, hen's eggs, and vegetable oils. In the case of milk, to improve its durability, some heating methods were applied, resulting in a further decrease in the light storage capacity of milk and consequently lead to a change in its components and quality, such as protein denaturation dephosphorylation and loss of vitamins. For this purpose, some pasteurized, homogenized, and ultra-high temperature milk samples with different fat percentages were prepared. The samples were exposed to white, red, and blue lights (light illumination). The UPE was mostly increased after light illumination was applied to these samples. It was revealed that the higher the light storage capacity, the lower the intensity of UPE. Based on these findings, a decrease and increase in the milk's light storage with low fat (1.5%) and natural fat content (3.5–3.8%) were noted, respectively. In the case of eggs, the aim was to find the origin of eggs while the egg yolk's desired color attracted the consumers. To identify the origin of eggs, 325 brown hens were divided into four different groups: soil, soil with free-range on vegetation, cage, and sand.

Further chemical analysis showed that no significant differences between the groups were evident. Although the eggs from cages or soil exhibited a lower UPE rate than free-range eggs (on vegetation and sand), it demonstrated that the source of UPE is not a single chemical substance. In edible oil, 24 different types of sunflower oils were subjected to UPE measurement in three groups (without external illumination, after white and red lights illumination). The quality of sunflower oils is characterized by three-factor values, including light storage capacity, decomposition procedures (e.g., low quality of storage and aging), and the order's value in the sense

of physiological and nutritional values. A high rate of UPE after red light illumination can be associated with decomposition procedures. The irradiation of food breaks chemical bonds and consequently forms radicals. By illumination with light, the electronically excited radicals fall to their base states, and due to these different energy levels, the light is emitted. Therefore, food irradiation shows a much higher emission rate (higher by a factor of 50) after light illumination than nonirradiated ones (Lambing, 1992).

Another study aimed to monitor the UPE accompanying autoxidation and water–biopolymer interactions in cereal products using the CCD technique. While the hydration of cellulose, dextran, or starch chains resulted in the hydrogen-bond formation and, consequently, accumulation of the excitation energy, incorporating water into cereal products enhanced the UPE (Slawinska and Slawinski, 1997, 1998).

The quality of tomato fruit was studied using UPE measurement. The fruits were harvested at four maturity stages (green–orange, orange–red, light red, and red) with almost the same size and weight. They were stored at a particular temperature (20 °C) and humidity (80% RH) for 10 days. It was found that the UPE was directly related to harvest maturity. It was mentioned that UPE could be used as a nondestructive method to evaluate tomato quality (Triglia *et al.*, 1998).

The UPE was measured for rice (*Oryza sativa* L.) seeds, which were stored during different years (1996, 1998, 1999, and 2001), and the correlation between the degree of aging of rice seeds and the intensity of UPE was noted. It was observed that the rice seeds stored for a shorter period had a stronger intensity of UPE in early imbibition. Moreover, a significant correlation was reported between the germination rates of rice seeds and the intensity of UPE (Chen *et al.*, 2003).

The coffee seed viability was studied by the aid of a UPE measurement. For this purpose, six coffee seeds were selected to measure UPE after exposure to white light at a constant temperature (22 °C). The germination rates were recorded on the 15th and 30th day. The proposed method with further investigations can promote advances in storing methods via the UPE technique (Galleg *et al.*, 2004).

The quality parameters of differently cultivated carrots from an organic farm and conventional farms in Austria were studied for 5 years (1998–2003) with different quality assessment methods, including sensor tests, food preference tests with laboratory rats, decomposition tests, P-value determination, chemical analysis, and UPE. In this work, the carrot slices were put under a light bulb,

Table 2. The relation between food quality and UPE.

Authors/years	Sample	Aim	Preparation	Conclusion
Lambing, 1992	Pasteurized, homogenized, and ultra-high-temperature milk	Assessing the quality of milk	The samples were exposed to white, red, and blue lights (light illumination)	The UPE was mostly increased after light illumination was applied to samples
Lambing, 1992	Hen's egg	Finding the origin of eggs	Three hundred twenty-five brown hens were divided into four different groups	The eggs from cages or soil exhibit a lower UPE rate than eggs from free-range eggs
Lambing, 1992	Sunflower oils	Assessing the quality of sunflower oils	Sunflower oils were divided into three groups	A high rate of UPE after red light illumination was observed
Slawinska and Slawinski, 1997, 1998	Cereal products	Evaluating the quality of cereal products	Hydration of cereal food products	The incorporation of water into cereal products enhanced the UPE intensity
Triglia et al., 1998	Tomato fruit	Studying the quality of tomato fruit	The tomato fruits were harvested at four maturity stages and stored for 10 days	The tomato fruits with similar colors had significant differences in the UPE intensity
Chen et al., 2003	Rice seeds	Finding the correlation between the degree of aging and the germination rates of rice seeds with the intensity of UPE	Rice seeds are stored in different years	It was revealed that the rice seeds stored for a shorter period had a more vigorous intensity of UPE in early imbibition.
Gallep et al., 2004	Coffee seeds	Finding the correlation between the coffee seeds viability and UPE intensity	Six groups of samples were exposed to white light at a constant temperature	The proposed method with further investigations can be employed to promote advances in storing methods via the UPE technique
Velimirov, 2005	Carrots	For the investigation of the quality parameters of cultivated carrots from organic and conventional farms	The same carrots cultivar grown in the same region were considered for 5 years	A significantly better capacity to store biophotons in organic carrot samples was noted while compared with conventional samples
Gallep and Dos Santos, 2007	Wheat seeds germinating	Investigating the relation between seedling growth and UPE intensity	The samples were placed in three different wastewater sediment solutions	A relation between the seedling growth and the detected light intensity was increased over time
Grashorn and Egerer, 2007	Organic and conventional eggs	To evaluate the quality of the parameters of the eggs	Four different groups of eggs were considered to measure the UPE intensity	The organic eggs showed a higher UPE with a slower decline than conventional ones
Wang and Yu, 2009	Wheat grain	Investigation of the correlation between UPE intensity and vigor of irradiated wheat grain and its irradiation dose	Wheat grain and wheat flour were irradiated by 60Co sources. samples were selected and stored for some months for UPE measurement	The UPE analysis cannot be used to detect the irradiation dose but is capable of determining vigor
Hossu et al., 2010	Sweet potato	Correlation between the UPE measurement and the quality of samples	Ag NP solution with different concentrations was added on the surface of the samples. Then UPE measurements were carried out for three cases	UPE was enhanced as much as 15 times by adding Ag NP
Kausar et al. 2012	soybean	The correlation between the UPE and seedling growth under flooding and drought stresses	The 3-day-old samples were exposed to flooding or drought stresses for 2, 4, and 6 days	Differential patterns of UPE were detected for considered days, and maximum UPE was evident under flooding stress
Khatoon et al. 2012	soybean	The correlation between seedling growth under flooding stress and UPE intensity	The 2-day-old samples were exposed to flooding or drought stresses for 5 days	The UPE was increased in flooded soybean samples compared to untreated samples

(Continued)

Table 2. Continued

Authors/years	Sample	Aim	Preparation	Conclusion
Komatsu <i>et al.</i> , 2014	soybean	Correlation between seedling growth under abiotic stresses and UPE intensity	The 2-day-old samples were exposed to abiotic stresses for 5 days	Differential patterns of UPE were detected for considered days, and maximum UPE was evident under flooding stress compared to drought stress; and also, the UPE in the leaves treated with cadmium was higher than untreated soybean
Kamal and Komatsu, 2016	Soybean	Investigating the molecular systems based on flooding stressed sample roots and UPE evaluation	Exposure of the sample roots to flooding stress along with light and dark situations	UPE was considerably increased by flooding stress, then decreased with a continued flooding exposure
Cordeiro <i>et al.</i> , 2017	Water samples	To evaluate the contamination of water samples from a river	The UPE of water samples from a river near Curitiba City in Brazil by coliform was assessed	The UPE measurements are an effective way to discriminate between contaminated and noncontaminated samples
Grasso <i>et al.</i> , 2018	Seeds of watermelon	Verification of the growth performance of inherently aged or damaged seeds of watermelon	The samples were placed in a controlled dark condition until a 2-mm root length was reached. To the spectral analysis of UPE, the interference filters were employed	The UPE measurements were strictly related to the biological state of the system under analysis
Nawara <i>et al.</i> , 2018	Food samples	Designed and manufacture the station to assess the quality of food	The designed device was to measure the degree of UPE of organic matters and processes	The measuring station is a useful way to compare traditional food quality with similar foods produced by industrial methods
Sun <i>et al.</i> , 2019	Herbal materials	To identify authentic from counterfeit herbal materials	For this purpose, four category tests were selected	UPE can be utilized as a fast, simple, and inexpensive method, rather than conventional techniques to identify herbal materials
Gałażka-Czarnecka <i>et al.</i> , 2019	Eggs	Assessing the quality of eggs	Two different hens, including the caged and free-range, were selected to determine their freshness and quality	It was observed that UPE from free-range eggs had eight times higher intensity than eggs from caged hens. They also found that the eggs from free-range hens had more than three times higher carotenoid than eggs from caged hens
Jia <i>et al.</i> , 2020	Herbal materials	To study the properties of aged and contemporary Chinese herbal materials	The UPE analysis was performed on some selected groups of aged and contemporary materials of their corresponding species	They found that the UPE technique can be employed to differentiate the aged and contemporary samples

UPE: ultra-weak photon emission; NP: nanoparticles.

and UPE was measured. A significantly better capacity to store biophotons in organic carrot samples was compared with conventional samples (Velimirov, 2005).

The photon-counting of wheat seeds germinating in three different wastewater sediment solutions was analyzed using a PMT device, and correlation with seedling development was studied. It was indicated that there was an increasing relation between seedling growth and detected light intensity over time (Gallego and Dos Santos, 2007).

The quality of organic and conventional eggs (for 1 year) was investigated using a UPE measurement. In this regard, four different forms of production systems (barn, cage, organic, and free-range) were considered to evaluate the quality of eggs based on conventional (egg mass, shell-breaking strength, albumen height, the proportion of yolk, fatty acid profile, and yellow color) quality criteria and UPE measurements. The results depicted higher UPE with a slower declining trend for organic eggs. It was reported that the measurement of UPE could be a suitable method for evaluating the quality of organic eggs (Grashorn and Egerer, 2007).

Correlation between UPE intensity and vigor of irradiated wheat grain and its irradiation dose was investigated. At first, wheat grain and wheat flour were irradiated by ^{60}Co sources with a dose rate of 1 kGy/h, including 0, 0.6, 1.5, 2.4, and 3 kGy. Samples were stored for 0, 6, 12, and 18 months under commercial storage conditions for UPE measurement. In summary, UPE analysis could not detect irradiation dose but was capable of determining vigor (Wang and Yu, 2009).

Hossu *et al.* (2010) tried to find a relation between the UPE and sweet potato samples' quality. They selected eight sweet potato roots with 5-mm thick disk slices and an average of nine samples from each root. The samples were placed in a Petri dish with 8-mL 3% sucrose (media). The samples were incubated for 1 week at a relative humidity of 90–95% and a temperature of 30 °C to increase storage quality. Different concentrations of the 2-mL solution of silver (Ag) nanoparticles (NP) were added on sweet potato samples' surfaces. Then UPE measurements were carried out for three cases (no media, media only, and adding Ag NP). They found that the UPE was enhanced by as much as 15 times by adding Ag NP. They mentioned that the UPE could provide useful information about the quality of biological material.

The relation between the UPE intensity and soybean seedling subjected to abiotic stresses was investigated. For this purpose, Kausar *et al.* (2012) (flooding and drought stresses), Khatoun *et al.* (2012) (flooding stress), and Komatsu *et al.* (2014) (flooding, drought, and

cadmium stresses) considered 2 or 3-day-old seedling samples and exposed to stresses above. They measured the activity of the APX and isoflavone reductase, which have related to the UPE measurement. They found that differential patterns of UPE were detected for considered days, and maximum UPE was evident under flooding stress compared to drought stress, and the UPE in the leaves treated with cadmium was higher than untreated soybean samples.

Flooding is abiotic stress that influences plant growth and crop yields. Kamal and Komatsu (2016) investigated the molecular systems based on flooding-stressed roots in soybean and UPE evaluation under light and dark conditions. They found that the UPE was considerably increased with light and dark conditions after flooding stress but decreased with continued flooding exposure. They also showed that increase in the activity of enzyme lysine ketoglutarate reductase/saccharopine dehydrogenase bifunctional was due to flooding stress, which consequently increased ROS rate scavenging and UPE.

The UPE measurement for evaluating microbial contamination (coliform group) of water samples from a river near Curitiba City in Brazil was assessed. It was observed that the UPE measurement is an effective way to discriminate between contaminated and noncontaminated water samples (Cordeiro *et al.*, 2017).

Grasso *et al.* (2018) verified the growth performance of inherently aged or damaged watermelon seeds by the UPE technique. For this purpose, they selected two lots of watermelon seeds, with 96 seeds in each lot. All the germination tests were performed using 12 dishes per lot, eight seeds in each dish, and a filter paper. To perform UPE measurements, the samples were placed in a controlled dark condition (at a temperature of 28.3°C) until a 2-mm root length was reached. For the spectral analysis of UPE, the interference filters (Edmund Optics; center wavelength 450, 550, and 650 nm, respectively) were employed. The results showed that the UPE measurements were strictly related to the system's biological state under analysis. They claimed that the proposed method could be used as a noninvasive and nondestructive technique for rapidly analyzing the seeds' viability and enhancing tools for seed-sorting systems.

Nawara *et al.* (2018) designed and manufactured a station to assess food quality. The designed device was to measure the degree of UPE of organic matters and processes. The manufactured instrument included a PC with counting, controlling systems, and measurements, including amplification and counting single photons (ESPC), control card, the light source for automatic recording of test results, and an application software created in the LabView environment. The measuring station

was a useful way to compare traditional food quality with similar foods produced by industrial methods. This device can also measure the quality parameters of food products.

Interesting research was conducted by Sun *et al.* (2019) for quality control of herbal materials. They employed the UPE for identifying authentic from counterfeit herbal materials. For this purpose, they used four-category tests, which included (i) authentic versus counterfeit materials; (ii) authentic versus adulterated materials; (iii) authentic versus sulfur-fumigated materials, and (iv) authentic versus dyed materials. The authors found that the UPE could be utilized as a fast, simple, and inexpensive method for identification of herbal materials in comparison to conventional techniques such as morphological and microscopic methods, chromatography and spectrum photometer analyses, molecular biology assays, and biomimetic technologies (Chen *et al.*, 2012; Xu *et al.*, 2015).

Recently, the traditional and UPE methods were used for determining egg quality. Measured quality parameters were the color at the La Roche point—YCF scale, pH, Haugh unit, and yolk color. Gałazka-Czarnecka *et al.* (2019) selected two different hens, including caged and free-range hens, with 60 eggs, yolk, and white, to determine the freshness and quality. Three yolks of each type of egg were mixed and considered as a sample. It was observed that UPE from eggs of free-range hens had eight times higher intensity than eggs from caged hens. The authors also found that the eggs of free-range hens had more than three times higher carotenoid than caged hens. They stated that further research is needed for using the UPE method as a food quality assessment tool.

Jia *et al.* (2020) studied the aged and contemporary properties of some Chinese herbal materials, including *Glycyrrhiza glabra* L., *Glycyrrhiza inflata* Batalin, *Glycyrrhiza uralensis* Fisch., *Curcuma aromatica* Salisb., *Zingiber officinale* Roscoe, *Acorus calamus* L., and *Alpinia officinarum* Hance. They implemented the UPE technique by PMT devices to differentiate the aged and contemporary samples. They found out that the UPE technique can be able to achieve the desired result. They also suggested that the UPE technique provides useful data on the storage time effect and the herbal medicines' quality assessment.

Conclusion

This review presents an overall framework on various aspects of UPE. Definition, detection techniques, and different applications of UPE were studied. The UPE results from the relaxation of the electronically excited species, resulting from oxidative metabolic and oxidative stresses.

Owing to the growing world population and food security, safety, and quality is imperative issues, the current study focuses on the applications of UPE on the subject of food quality. The proposed method with further investigations could open new horizons on many branches of sciences. This technique is non-invasive and nondestructive and a cheap, rapid, and real-time technique that does not need complex instrumentation.

References

- Abouzari, A. and Fakheri, B.A., 2015. Reactive oxygen species: generation, oxidative damage, and signal transduction. *International Journal of Life Sciences* 9(5): 3–17. <https://doi.org/10.3126/ijls.v9i5.12699>
- Adam, W., Kazakov, D.V. and Kazakov, V.P., 2005. Singlet-oxygen chemiluminescence in peroxide reactions. *Chemical Reviews* 105(9): 3371–3387. <https://doi.org/10.1021/cr0300035>
- Ahamed, G.J., Xu, W., Liu, A. and Chen, S., 2019. Endogenous melatonin deficiency aggravates high temperature-induced oxidative stress in *Solanum lycopersicum* L. *Environmental and Experimental Botany* 161: 303–311. <https://doi.org/10.1016/j.envexpbot.2018.06.006>
- Ahmad, P. (Ed.), 2013. *Oxidative damage to plants—antioxidant networks and signaling*. Elsevier, San Diego, CA.
- Bailey-Serres, J. and Mittler, R., 2006. The roles of reactive oxygen species in plant cells. *Plant Physiology* 141: 311. <https://doi.org/10.1104/pp.104.900191>
- Battin, E.E. and Brumaghim, J.L., 2009. Antioxidant activity of sulfur and selenium: a review of reactive oxygen species scavenging, glutathione peroxidase, and metal-binding antioxidant mechanisms. *Cell Biochemistry and Biophysics* 55(1): 1–23. <https://doi.org/10.1007/s12013-009-9054-7>
- Bennett, M., Mehta, M. and Grant, M., 2005. Biophoton imaging: a nondestructive method for assaying R gene responses. *Molecular Plant-Microbe Interactions* 18(2): 95–102. <https://doi.org/10.1094/MPMI-18-0095>
- Bischof, M., 2003. Introduction to integrative biophysics. In: *Integrative biophysics*. Springer, Dordrecht, Netherlands, pp. 1–115.
- Bischof, M., 2005. Biophotons—the light in our cells. *Journal of Optometric Phototherapy*, pp. 1–5.
- Burgos, R.C.R., Schoeman, J.C., van Winden, L.J., Červinková, K., Ramautar, R., Van Wijk, E.P., Cifra, M., Berger, R., Hankemeier, T. and van der Greef, J., 2017. Ultra-weak photon emission as a dynamic tool for monitoring oxidative stress metabolism. *Scientific Reports* 7(1):1–9. <https://doi.org/10.1038/s41598-017-01229-x>
- Caverzan, A., Piasecki, C., Chavarria, G., Stewart, C.N. and Vargas, L., 2019. Defenses against ROS in crops and weeds: the effects of interference and herbicides. *International Journal of Molecular Sciences* 20(5): 1086. <https://doi.org/10.3390/ijms20051086>
- Cheeseman, J., 2016. Food security in the face of salinity, drought, climate change, and population growth. In: *Halophytes for food security in dry lands*. Academic Press, San Diego, USA, pp. 111–123.

- Chen, S., Guo, B., Zhang, G., Yan, Z., Luo, G., Sun, S., Wu, H., Huang, L., Pang, X. and Chen, J., 2012. Advances of studies on new technology and method for identifying traditional Chinese medicinal materials. *Zhongguo Zhong yao za zhi= Zhongguo zhongyao zazhi= China Journal of Chinese Materia Medica* 37(8): 1043–1055. <https://doi.org/10.4268/CJCM20120801>
- Chen, W., Xing, D., Wang, J. and He, Y., 2003. Rapid determination of rice seed vigour by spontaneous chemiluminescence and singlet oxygen generation during early imbibition. *Luminescence: The Journal of Biological and Chemical Luminescence* 18(1): 19–24. <https://doi.org/10.1002/bio.695>
- Cifra, M., Fields, J.Z. and Farhadi, A., 2011. Electromagnetic cellular interactions. *Progress in Biophysics and Molecular Biology* 105(3): 223–246. <https://doi.org/10.1016/j.pbiomolbio.2010.07.003>
- Cifra, M. and Pospíšil, P., 2014. Ultra-weak photon emission from biological samples: definition, mechanisms, properties, detection and applications. *Journal of Photochemistry and Photobiology B: Biology* 139: 2–10. <https://doi.org/10.1016/j.jphotobiol.2014.02.009>
- Cordeiro, A., Fabris, J., Couto, G., Kalinowski, H. and Bertogna, E., 2017. Water assessment using ultra-weak bioluminescence. *Journal of Photochemistry and Photobiology B: Biology* 177: 39–43. <https://doi.org/10.1016/j.jphotobiol.2017.10.014>
- de Mello Gallep, C. and Robert, D., 2020. Time-resolved ultra-weak photon emission as germination performance indicator in single seedlings. *Journal of photochemistry and photobiology 1: 100001*. <https://doi.org/10.1016/j.jpap.2020.100001>
- Egawa, M., Kohno, Y. and Kumano, Y., 1999. Oxidative effects of cigarette smoke on the human skin. *International Journal of Cosmetic Science* 21(2): 83–98. <https://doi.org/10.1046/j.1467-2494.1999.181656.x>
- El-Mesery, H.S., Mao, H. and Abomohra, A.E.-F., 2019. Applications of nondestructive technologies for agricultural and food products quality inspection. *Sensors* 19(4): 846. <https://doi.org/10.3390/s19040846>
- Esmailpour, T., Fereydouni, E., Dehghani, F., Bókkon, I., Panjeshahin, M.-R., Császár-Nagy, N., Ranjbar, M. and Salari, V., 2020. An experimental investigation of ultraweak photon emission from adult murine neural stem cells. *Scientific Reports* 10: 1–13. <https://doi.org/10.1038/s41598-019-57352-4>
- Evelson, P., Ordóñez, C.P., Llesuy, S. and Boveris, A., 1997. Oxidative stress and in vivo chemiluminescence in mouse skin exposed to UVA radiation. *Journal of Photochemistry and Photobiology B: Biology* 38(2–3): 215–219. [https://doi.org/10.1016/S1011-1344\(96\)07437-4](https://doi.org/10.1016/S1011-1344(96)07437-4)
- Fedorova, G.F., Trofimov, A.V., Vasil'ev, R.F. and Vepintsev, T.L., 2007. Peroxy-radical-mediated chemiluminescence: mechanistic diversity and fundamentals for antioxidant assay. *Arkhivoc* 8, 163–215. <https://doi.org/10.3998/ark.5550190.0008.815>
- Gałązka-Czarnecka, I., Korzeniewska, E., Czarnecki, A., Sójka, M., Kielbasa, P. and Drózdź, T., 2019. Evaluation of quality of eggs from hens kept in caged and free-range systems using traditional methods and ultra-weak luminescence. *Applied Sciences* 9(12): 2430. <https://doi.org/10.3390/app9122430>
- Gallep, C., Conforti, E., Braghini, M., Maluf, M., Yan, Y. and Popp, F., 2004. Ultra-weak delayed luminescence in coffee seeds (*Coffea arabica* and *C. canephora*) and their germination potential: some indications for a photonic approach in seed viability. *Proceedings of 11th Brazilian Symposium of Microwave and Optoelectronics, São Paulo, Brazil, section p1–12*.
- Gallep, C. and Dos Santos, S., 2007. Photon-counts during germination of wheat (*Triticum aestivum*) in wastewater sediment solutions correlated with seedling growth. *Seed Science and Technology* 35(3): 607–614. <https://doi.org/10.15258/sst.2007.35.3.08>
- Gill, S.S. and Tuteja, N., 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry* 48(12): 909–930. <https://doi.org/10.1016/j.plaphy.2010.08.016>
- Giovenzana, V., Beghi, R., Civelli, R., Trapani, S., Migliorini, M., Cini, E., Zanoni, B. and Guidetti, R., 2017. Rapid determination of crucial parameters for the optimization of milling process by using visible/near infrared spectroscopy on intact olives and olive paste. *Italian Journal of Food Science* 29(2): 357–369. <https://doi.org/10.14674/1120-1770/ijfs.v560>
- Godfray, H.C.J., Beddington, J.R., Crute, I.R., Haddad, L., Lawrence, D., Muir, J.F., Pretty, J., Robinson, S., Thomas, S.M. and Toulmin, C., 2010. Food security: the challenge of feeding 9 billion people. *Science* 327(5967): 812–818. <https://doi.org/10.1126/science.1185383>
- Grashorn, M.A. and Egerer, U., 2007. Integrated assessment of quality of chicken organic eggs by measurement of dark luminescence. *Polish Journal of Food and Nutrition Sciences* 57(4 [A]): 191–194.
- Grasso, R., Gulino, M., Giuffrida, F., Agnello, M., Musumeci, F. and Scordino, A., 2018. Nondestructive evaluation of watermelon seeds germination by using delayed luminescence. *Journal of Photochemistry and Photobiology B: Biology* 187: 126–130. <https://doi.org/10.1016/j.jphotobiol.2018.08.012>
- Gu, Q., 1999. On coherence theory of biophoton emission. *Journal of the GCPD eV* 5(17): 17–20.
- Guo, J., Zhu, G., Li, L., Liu, H. and Liang, S., 2017. Ultraweak photon emission in strawberry fruit during ripening and aging is related to energy level. *Open Life Sciences* 12(1): 393–398. <https://doi.org/10.1515/biol-2017-0046>
- Guo, Y. and Tan, J., 2013. A biophotonic sensing method for plant drought stress. *Sensors and Actuators B: Chemical* 188: 519–524. <https://doi.org/10.1016/j.snb.2013.07.020>
- Gupta, D.K., Palma, J.M. and Corpas, F.J., 2015. Reactive oxygen species and oxidative damage in plants under stress. Springer. Cham, Switzerland.
- Hossu, M., Ma, L. and Chen, W., 2010. Nonlinear enhancement of spontaneous biophoton emission of sweet potato by silver nanoparticles. *Journal of Photochemistry and Photobiology B: Biology* 99: 44–48. <https://doi.org/10.1016/j.jphotobiol.2010.02.002>
- Inagaki, H., Imaizumi, T., Wang, G.-X., Tominaga, T., Kato, K., Iyozumi, H. and Nukui, H., 2007. Spontaneous ultraweak photon emission from rice (*Oryza sativa* L.) and paddy weeds treated with a sulfonylurea herbicide. *Pesticide Biochemistry and Physiology* 89(2): 158–162. <https://doi.org/10.1016/j.pestbp.2007.05.005>

- Inagaki, H., Imaizumi, T., Wang, G.-X., Tominaga, T., Kato, K., Iyozumi, H. and Nukui, H., 2009. Sulfonyl urea-resistant biotypes of *Monochoria vaginalis* generate higher ultraweak photon emissions than the susceptible ones. *Pesticide Biochemistry and Physiology* 95(3): 117–120. <https://doi.org/10.1016/j.pestbp.2009.08.002>
- Inagaki, H., Ishida, Y., Uchino, A., Kato, K., Kageyama, C., Iyozumi, H. and Nukui, H., 2008. Difference in ultraweak photon emissions between sulfonylurea-resistant and sulfonylurea-susceptible biotypes of *Scirpus juncooides* following the application of a sulfonylurea herbicide. *Weed Biology and Management* 8(2): 78–84. <https://doi.org/10.1111/j.1445-6664.2008.00278.x>
- Iyozumi, H., Kato, K. and Makino, T., 2002. Spectral shift of ultraweak photon emission from sweet potato during a defense response. *Photochemistry and Photobiology* 75(3): 322–325. [https://doi.org/10.1562/0031-8655\(2002\)075%3C0322:SSOUPE%3E2.0.CO;2](https://doi.org/10.1562/0031-8655(2002)075%3C0322:SSOUPE%3E2.0.CO;2)
- Janků, M., Luhová, L. and Petřiválský, M., 2019. On the origin and fate of reactive oxygen species in plant cell compartments. *Antioxidants* 8(4): 105. <https://doi.org/10.3390/antiox8040105>
- Jia, Y., Sun, M., Shi, Y., Zhu, Z., van Wijk, E., van Wijk, R., van Andel, T. and Wang, M., 2020. A comparative study of aged and contemporary Chinese herbal materials by using delayed luminescence technique. *Chinese medicine* 15: 6. <https://doi.org/10.1186/s13020-020-0287-0>
- Kageyama, C., Kato, K., Iyozumi, H., Inagaki, H., Yamaguchi, A., Furuse, K. and Baba, K., 2006. Photon emissions from rice cells elicited by N-acetylchitoooligosaccharide are generated through phospholipid signaling in close association with the production of reactive oxygen species. *Plant Physiology and Biochemistry* 44(11–12): 901–909. <https://doi.org/10.1016/j.plaphy.2006.09.010>
- Kamal, A.H.M. and Komatsu, S., 2016. Proteins involved in biophoton emission and flooding-stress responses in soybean under light and dark conditions. *Molecular Biology Reports* 43(2): 73–89. <https://doi.org/10.1007/s11033-015-3940-4>
- Karuppanapandian, T., Moon, J.-C., Kim, C., Manoharan, K. and Kim, W., 2011. Reactive oxygen species in plants: their generation, signal transduction, and scavenging mechanisms. *Australian Journal of Crop Science* 5(6): 709.
- Kato, K., Iyozumi, H., Kageyama, C., Inagaki, H., Yamaguchi, A. and Nukui, H., 2014. Application of ultra-weak photon emission measurements in agriculture. *Journal of Photochemistry and Photobiology B: Biology* 139: 54–62. <https://doi.org/10.1016/j.jphotobiol.2014.06.010>
- Kausar, R., Hossain, Z., Makino, T. and Komatsu, S., 2012. Characterization of ascorbate peroxidase in soybean under flooding and drought stresses. *Molecular Biology Reports* 39(12): 10573–10579. <https://doi.org/10.1007/s11033-012-1945-9>
- Kehrer, J.P., 2000. The Haber–Weiss reaction and mechanisms of toxicity. *Toxicology* 149(1): 43–50. [https://doi.org/10.1016/S0300-483X\(00\)00231-6](https://doi.org/10.1016/S0300-483X(00)00231-6)
- Khatoun, A., Rehman, S., Hiraga, S., Makino, T. and Komatsu, S., 2012. Organ-specific proteomics analysis for identification of response mechanism in soybean seedlings under flooding stress. *Journal of Proteomics* 75(18): 5706–5723. <https://doi.org/10.1016/j.jprot.2012.07.031>
- Khairilipour, K., Ahmadi, H., Rajabipour, A., Rafiee, S., Javan-Nikkhah, M., Jayas, D. and Siliveru, K., 2016. Detection of fungal infection in pistachio kernel by long-wave near-infrared hyperspectral imaging technique. *Quality Assurance and Safety of Crops and Foods* 8(1): 129–135. <https://doi.org/10.3920/QAS2015.0606>
- Kobayashi, M., Sasaki, K., Enomoto, M. and Ehara, Y., 2006. Highly sensitive determination of transient generation of biophotons during hypersensitive response to cucumber mosaic virus in cowpea. *Journal of Experimental Botany* 58(3): 465–472. <https://doi.org/10.1093/jxb/erl215>
- Komatsu, S., Kamal, A.H.M., Makino, T. and Hossain, Z., 2014. Ultraweak photon emission and proteomics analyses in soybean under abiotic stress. *Biochimica et Biophysica Acta (BBA)—Proteins and Proteomics* 1844(7): 1208–1218.
- Krieger-Liszczay, A., 2005. Singlet oxygen production in photosynthesis. *Journal of Experimental Botany* 56(411): 337–346.
- Lambing, K., 1992. Biophoton measurement as a supplement to the conventional consideration of food quality. In: Popp, F. A., Li, K. H. and Gu, Q. (eds) *Recent advances in biophoton research and its applications*. World Scientific, Singapore, Singapore, pp. 393–413.
- Liang, X., Wang, Z., Gao, M., Wu, S., Zhang, J., Liu, Q., Yu, Y., Wang, J. and Liu, W., 2019. Cyclic stretch-induced oxidative stress by mitochondrial and NADPH oxidase in retinal pigment epithelial cells. *BMC Ophthalmology* 19(1): 79. <https://doi.org/10.1186/s12886-019-1087-0>
- Madl, P., 2014. Detection and measurement of biogenic ultra-weak photon emission. In: Fels, D., Cifra, M. and Scholkmann, F. (eds.) *Field of the cells*. Research Signpost, Trivandrum, Kerala, India. pp. 55–70.
- Magwaza, L.S., Ford, H.D., Cronje, P.J., Opara, U.L., Landahl, S., Tatam, R.P. and Terry, L.A., 2013. Application of optical coherence tomography to non-destructively characterise rind breakdown disorder of ‘Nules Clementine’ mandarins. *Postharvest Biology and Technology* 84: 16–21. <https://doi.org/10.1016/j.postharvbio.2013.03.019>
- Makino, T., Kato, K., Iyozumi, H., Honzawa, H., Tachiiri, Y. and Hiramatsu, M., 1996. Ultraweak luminescence generated by sweet potato and *Fusarium oxysporum* interactions associated with a defense response. *Photochemistry and Photobiology* 64(6): 953–956. <https://doi.org/10.1111/j.1751-1097.1996.tb01860.x>
- Mansfield, J.W., 2005. Biophoton distress flares signal the onset of the hypersensitive reaction. *Trends in Plant Science* 10(7): 307–309. <https://doi.org/10.1016/j.tplants.2005.05.007>
- Mayorga Burrezo, P., Jiménez, V.G., Blasi, D., Ratera, I., Campaña, A.G. and Veciana, J., 2019. Organic free radicals as circularly polarized luminescence emitters. *Angewandte Chemie (International Edition)* 58(45): 16282–16288. <https://doi.org/10.1002/anie.201909398>
- McCaig, C.D., Rajnicek, A.M., Song, B. and Zhao, M., 2005. Controlling cell behavior electrically: current views and future potential. *Physiological Reviews* 85(3): 943–978. <https://doi.org/10.1152/physrev.00020.2004>
- McCarthy, U., Uysal, I., Melis, R.B., Mercier, S., Donnell, C.O. and Ktenioudaki, A., 2018. Global food security—issues, challenges

- and technological solutions. *Trends in Food Science and Technology* 77: 11–20. <https://doi.org/10.1016/j.tifs.2018.05.002>
- Montillet, J.-L., Chamnongpol, S., Rustérucci, C., Dat, J., Van De Cotte, B., Agnel, J.-P., Battesti, C., Inzé, D., Van Breusegem, F. and Triantaphylides, C., 2005. Fatty acid hydroperoxides and H₂O₂ in the execution of hypersensitive cell death in tobacco leaves. *Plant Physiology* 138(3): 1516–1526. <https://doi.org/10.1104/pp.105.059907>
- Moraes, T.A., Barlow, P.W., Klingelé, E. and Gallep, C.M., 2012. Spontaneous ultra-weak light emissions from wheat seedlings are rhythmic and synchronized with the time profile of the local gravimetric tide. *Naturwissenschaften* 99(6): 465–472. <https://doi.org/10.1007/s00114-012-0921-5>
- Mousavi Khaneghah, A., 2016. An overview on some of important sources of natural antioxidants. *International Food Research Journal* 23(3): 928–933.
- Münzel, T., Sørensen, M., Schmidt, F., Schmidt, E., Steven, S., Kröller-Schön, S. and Daiber, A., 2018. The adverse effects of environmental noise exposure on oxidative stress and cardiovascular risk. *Antioxidants and Redox Signaling* 28(9): 873–908. <https://doi.org/10.1089/ars.2017.7118>
- Nakashima, Y., Ohta, S. and Wolf, A.M., 2017. Blue light-induced oxidative stress in live skin. *Free Radical Biology and Medicine* 108: 300–310. <https://doi.org/10.1016/j.freeradbiomed.2017.03.010>
- Nawara, P., Gliniak, M., Popardowski, E., Szczuka, M. and Trzyniec, K., 2018. Control system of a prototype measurement system for the identification of ultra-low photonic emission of organic materials. *Progress in Applied Electrical Engineering (PAEE)*, June 18–22, 2018. Koscielisko, Poland, pp. 35–37.
- Nukui, H., Inagaki, H., Iyozumi, H. and Kato, K., 2013. Biophoton emissions in sulfonylurea herbicide-resistant weeds. In: *Herbicides—advances in research*, InTech Open Science, Rijeka, Croatia, pp. 219–235. <http://dx.doi.org/10.5772/55846>
- Ohya, T., Kurashige, H., Okabe, H. and Kai, S., 2000. Early detection of salt stress damage by biophotons in red bean seedling. *Japanese Journal of Applied Physics* 39(6R): 3696. <https://doi.org/10.1143/JJAP.39.3696>
- Ohya, T., Yoshida, S., Kawabata, R., Okabe, H. and Kai, S., 2002. Biophoton emission due to drought injury in red beans: possibility of early detection of drought injury. *Japanese Journal of Applied Physics* 41(7R): 4766. <https://doi.org/10.1143/JJAP.41.4766>
- Omar, A. and MatJafri, M., 2013. Principles, methodologies and technologies of fresh fruit quality assurance. *Quality Assurance and Safety of Crops and Foods* 5(3): 257–271. <https://doi.org/10.3920/QAS2012.0175>
- Ortega-Ojeda, E., Calcerrada, M., Ferrero, A., Campos, J. and Garcia-Ruiz, C., 2018. Measuring the human ultra-weak photon emission distribution using an electron-multiplying, charge-coupled device as a sensor. *Sensors* 18(4): 1152. <https://doi.org/10.3390/s18041152>
- Pitzschke, A., Forzani, C. and Hirt, H., 2006. Reactive oxygen species signaling in plants. *Antioxidants and Redox Signaling* 8(9–10): 1757–1764. <https://doi.org/10.1089/ars.2006.8.1757>
- Pospíšil, P., Prasad, A. and Rác, M., 2014. Role of reactive oxygen species in ultra-weak photon emission in biological systems. *Journal of Photochemistry and Photobiology B: Biology* 139: 11–23. <https://doi.org/10.1016/j.jphotobiol.2014.02.008>
- Pospíšil, P., Prasad, A. and Rác, M., 2019. Mechanism of the formation of electronically excited species by oxidative metabolic processes: role of reactive oxygen species. *Biomolecules* 9(7): 258. <https://doi.org/10.3390/biom9070258>
- Prasad, A., Gouripeddi, P., Devireddy, H.R.N., Ovsii, A., Rachakonda, D.P., Wijk, R.V. and Pospíšil, P., 2020. Spectral Distribution of Ultra-Weak Photon Emission as a Response to Wounding in Plants: An In Vivo Study. *Biology* 9: 139. <https://doi.org/10.3390/biology9060139>
- Prasad, A. and Pospíšil, P., 2011. Linoleic acid-induced ultra-weak photon emission from *Chlamydomonas reinhardtii* as a tool for monitoring of lipid peroxidation in the cell membranes. *PLoS One* 6(7): 1–10. <https://doi.org/10.1371/journal.pone.0022345>
- Prasad, A., Rossi, C., Lamponi, S., Pospíšil, P. and Foletti, A., 2014. New perspective in cell communication: Potential role of ultra-weak photon emission. *Journal of Photochemistry and Photobiology B: Biology* 139: 47–53. <https://doi.org/10.1016/j.jphotobiol.2014.03.004>
- Prosekov, A.Y. and Ivanova, S.A., 2018. Food security: the challenge of the present. *Geoforum* 91: 73–77. <https://doi.org/10.1016/j.geoforum.2018.02.030>
- Racchi, M.L., 2013. Antioxidant defenses in plants with attention to *Prunus* and *Citrus* spp. *Antioxidants* 2(4): 340–369. <https://doi.org/10.3390/antiox2040340>
- Rafii-Tabar, H. and Rafieiolhosseini, N., 2015. Different aspects of ultra-weak photon emissions: a review article. *Iranian Journal of Medical Physics* 12(3): 137–144.
- Rahnama, M., Tuszynski, J.A., Bokkon, I., Cifra, M., Sardar, P. and Salari, V., 2011. Emission of mitochondrial biophotons and their effect on electrical activity of membrane via microtubules. *Journal of Integrative Neuroscience* 10(1): 65–88. <https://doi.org/10.1142/S0219635211002622>
- Rastogi, A. and Pospíšil, P., 2010. Effect of exogenous hydrogen peroxide on biophoton emission from radish root cells. *Plant physiology and biochemistry* 48: 117–123. <https://doi.org/10.1016/j.plaphy.2009.12.01>
- Rastogi, A. and Pospíšil, P., 2011. Spontaneous ultraweak photon emission imaging of oxidative metabolic processes in human skin: effect of molecular oxygen and antioxidant defense system. *Journal of Biomedical Optics* 16(9): 096005. <https://doi.org/10.1117/1.3616135>
- Rastogi, A. and Pospíšil, P., 2012. Production of hydrogen peroxide and hydroxyl radical in potato tuber during the necrotrophic phase of hemibiotrophic pathogen *Phytophthora infestans* infection. *Journal of Photochemistry and Photobiology B: Biology* 117: 202–206. <https://doi.org/10.1016/j.jphotobiol.2013.03.012>
- Saeidfirozeh, H., Shafiekhani, A., Cifra, M. and Masoudi, A.A., 2018. Endogenous chemiluminescence from germinating *arabidopsis thaliana* seeds. *Scientific Reports* 8(1): 1–10. <https://doi.org/10.1038/s41598-018-34485-6>
- Saikolappan, S., Kumar, B., Shishodia, G., Koul, S. and Koul, H.K., 2019. Reactive oxygen species and cancer: a complex interaction. *Cancer Letters* 452: 132–143. <https://doi.org/10.1016/j.canlet.2019.03.020>

- Salari, V., Tuszyński, J., Bokkon, I., Rahnama, M. and Cifra, M., 2011. On the photonic cellular interaction and the electric activity of neurons in the human brain. *Journal of Physics: Conference Series* 329:1–9. <https://doi.org/10.1088/1742-6596/329/1/012006>
- Salari, V., Valian, H., Bassereh, H., Bókkon, I. and Barkhordari, A., 2015. Ultraweak photon emission in the brain. *Journal of Integrative Neuroscience* 14(3): 419–429. <https://doi.org/10.1142/S0219635215300012>
- Saueremann, G., Mei, W.P., Hoppe, U. and Stáb, F., 1999. Ultraweak photon emission of human skin *in vivo*: influence of topically applied antioxidants on human skin. In: Packer, L. (ed.) *Methods in enzymology*, Vol. 300. Elsevier, Amsterdam, the Netherlands, pp. 419–428.
- Schinabeck, T.-M., Weigler, F., Flöter, E. and Mellmann, J., 2018. Variability in determination of the single kernel moisture content of grain by means of TD-NMR spectroscopy. *Quality Assurance and Safety of Crops and Foods* 10(1): 75–82. <https://doi.org/10.3920/QAS2017.1149>
- Scholkmann, F., Fels, D. and Cifra, M., 2013. Non-chemical and non-contact cell-to-cell communication: a short review. *American Journal of Translational Research* 5(6): 586.
- Shanei, A., Alinasab, Z., Kiani, A. and Nematollahi, M., 2017. Detection of ultraweak photon emission (UPE) from cells as a tool for pathological studies. *Journal of Biomedical Physics and Engineering* 7(4): 389.
- Sies, H., Berndt, C. and Jones, D.P., 2017. Oxidative stress. *Annual Review of Biochemistry* 86: 715–748. <https://doi.org/10.1146/annurev-biochem-061516-045037>
- Singh, A., Yashavardhan, M., Kalita, B., Ranjan, R., Bajaj, S., Prakash, H. and Gupta, M.L., 2017. Podophyllotoxin and rutin modulates ionizing radiation-induced oxidative stress and apoptotic cell death in mice bone marrow and spleen. *Frontiers in Immunology* 8: 183. <https://doi.org/10.3389/fimmu.2017.00183>
- Slawinska, D. and Slawinski, J., 1997. Chemiluminescence of cereal products I. Kinetics, activation energy and effect of solvents. *Journal of Bioluminescence and Chemiluminescence* 12(5): 249–259. [https://doi.org/10.1002/\(SICI\)1099-1271\(199709/10\)12:5%3C249::AID-BIO453%3E3.0.CO;2-X](https://doi.org/10.1002/(SICI)1099-1271(199709/10)12:5%3C249::AID-BIO453%3E3.0.CO;2-X)
- Slawinska, D. and Sławinski, J., 1998. Chemiluminescence of cereal products III. Two-dimensional photocount imaging of chemiluminescence. *Journal of Bioluminescence and Chemiluminescence* 13(1): 21–24. [https://doi.org/10.1002/\(SICI\)1099-1271\(199801/02\)13:1%3C21::AID-IO462%3E3.0.CO;2-H](https://doi.org/10.1002/(SICI)1099-1271(199801/02)13:1%3C21::AID-IO462%3E3.0.CO;2-H)
- Sun, M., Wang, S., Jing, Y., Li, L., He, M., Jia, Y., van Wijk, E., Wang, Y., Wang, Z. and Wang, M., 2019. Application of delayed luminescence measurements for the identification of herbal materials: a step toward rapid quality control. *Chinese Medicine* 14(1): 1–13. <https://doi.org/10.1186/s13020-019-0269-2>
- Thannickal, V.J. and Fanburg, B.L., 2000. Reactive oxygen species in cell signaling. *American Journal of Physiology—Lung Cellular and Molecular Physiology* 279(6): L1005–L1028. <https://doi.org/10.1152/ajplung.2000.279.6.L1005>
- Triglia, A., La Malfa, G., Musumeci, F., Leonardi, C. and Scordino, A., 1998. Delayed luminescence as an indicator of tomato fruit quality. *Journal of Food Science* 63(3): 512–515. <https://doi.org/10.1111/j.1365-2621.1998.tb15775.x>
- Tripathy, B.C. and Oelmüller, R., 2012. Reactive oxygen species generation and signaling in plants. *Plant Signaling and Behavior* 7(12): 1621–1633. <https://doi.org/10.4161/psb.22455>
- Tsuchida, K., Iwasa, T. and Kobayashi, M., 2019. Imaging of ultraweak photon emission for evaluating the oxidative stress of human skin. *Journal of Photochemistry and Photobiology B: Biology* 198: 111562. <https://doi.org/10.1016/j.jphotobiol.2019.111562>
- Tsugane, K., Kobayashi, K., Niwa, Y., Ohba, Y., Wada, K. and Kobayashi, H., 1999. A recessive Arabidopsis mutant that grows photoautotrophically under salt stress shows enhanced active oxygen detoxification. *The Plant Cell* 11(7): 1195–1206. <https://doi.org/10.1105/tpc.11.7.1195>
- Van Wijk, R., 2001. Bio-photons and bio-communication. *Journal of Scientific Exploration* 15(2): 183–197.
- Van Wijk, E.P., Van Wijk, R. and Bosman, S., 2010. Using ultraweak photon emission to determine the effect of oligomeric proanthocyanidins on oxidative stress of human skin. *Journal of Photochemistry and Photobiology B: Biology* 98(3): 199–206. <https://doi.org/10.1016/j.jphotobiol.2010.01.003>
- Velimirov, A., 2005. The consistently superior quality of carrots from one organic farm in Austria compared with conventional farms. *Proceedings of the 15th IFOAM Organic World Congress “Researching and Shaping Sustainable Systems”, September 21–23, 2005, Adelaide*, pp. 1–4.
- Wang, J. and Yu, Y., 2009. Relationship between ultra-weak bioluminescence and vigour or irradiation dose of irradiated wheat. *Luminescence: The Journal of Biological and Chemical Luminescence* 24(4): 209–212. <https://doi.org/10.1002/bio.1096>
- Xu, Y., Song, W., Zhou, P., Li, P. and Li, H., 2015. Morphological and microscopic characterization of five commonly used testacean traditional Chinese medicines. *Acta Pharmaceutica Sinica B* 5(4): 358–366. <https://doi.org/10.1016/j.apsb.2015.03.014>
- Yang, M., Ding, W., Liu, Y., Fan, H., Bajpai, R.P., Fu, J., Pang, J., Zhao, X. and Han, J., 2017. Ultra-weak photon emission in healthy subjects and patients with type 2 diabetes: evidence for a non-invasive diagnostic tool. *Photochemical and Photobiological Sciences* 16(5): 736–743. <https://doi.org/10.1039/C6PP00431H>
- Yang, M., Pang, J., Liu, J., Liu, Y., Fan, H. and Han, J., 2015. Spectral discrimination between healthy people and cold patients using spontaneous photon emission. *Biomedical Optics Express* 6(4): 1331–1339. <https://doi.org/10.1364/BOE.6.001331>
- Yoshinaga, N., Kato, K., Kageyama, C., Fujisaki, K., Nishida, R. and Mori, N., 2006. Ultraweak photon emission from herbivory-injured maize plants. *Naturwissenschaften* 93(1): 38–41. <https://doi.org/10.1007/s00114-005-0059-9>
- Zhong, J. and Wang, X., 2019. An introduction to evaluation technologies for food quality. In: *Evaluation technologies for food quality*. Woodhead Publishing, Duxford, UK, pp. 1–3.