

## Gastrodin provides neuroprotection in models of traumatic brain injury via Nrf2 signaling pathway

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### Abstract

Gastrodin is one of the main active components of *Gastrodia elata* and has significant therapeutic value for various nervous system diseases. Its medicinal properties include smooth muscle relaxation, anti-necrosis, anti-aging, and anti-apoptosis effects. However, its possible effects on traumatic brain injury (TBI) are still unclear. In this study, the effects of gastrodin on TBI rats were investigated. The results proved that gastrodin had neuroprotective effect on TBI through alleviating brain deficits, decreasing brain water content, inhibiting neuronal apoptosis, and suppressing oxidative stress in brain tissues of TBI rats. Mechanically, gastrodin upregulated the expression of Nrf2 downstream proteins, suggesting the activation of Nrf2 pathway in brain tissues of TBI rats. In conclusion, gastrodin provided neuroprotection in TBI rats via Nrf2 pathway.

**Keywords:** apoptosis; gastrodin; Nrf2 pathway; oxidative stress; traumatic brain injury (TBI)

### Introduction

Traumatic brain injury (TBI) is a common injury in both peacetime and wartime (Schmitt *et al.*, 2021). It is estimated that there are about 10 million TBI patients worldwide each year, with a high prevalence of mild TBI (mTBI), accounting for about 76%–83%. The pathophysiological mechanisms of TBI are excitatory toxic injury, inflammation, increased vascular permeability, and oxidative stress (Suntai *et al.*, 2021). The series of pathological changes further aggravate the secondary brain damage caused by TBI, which is characterized by oxidative stress and inflammatory processes (Huibregtse *et al.*, 2021). Depletion of antioxidant enzymes could lead to excessive production of reactive oxygen species (ROS), which might destroy cellular components such as protein, lipid, and DNA, thus exacerbating neuronal apoptosis (Thomas *et al.*, 2021). Brain dysfunction after TBI is not only due to the original external mechanical forces, such as the role of primary injury, but also to a large extent related to the “secondary attack” of neurons

complicated by the injury, called secondary neuronal injury, whose mechanisms mainly include calcium overload, excitability amino acid toxic effects, mitochondrial dysfunction, oxidative stress, etc. Therefore, it is very important to find an effective method to reduce the oxidative stress caused by TBI.

Gastrodin is one of the main active components of *Gastrodia elata*, which has been isolated and confirmed as a traditional Chinese herbal medicine (Qin *et al.*, 2021). Gastrodin is the main bioactive ingredient from the rhizome of *Gastrodia elata*, and has significant therapeutic value for nervous system diseases (Yao *et al.*, 2019). In addition, its medicinal properties include smooth muscle relaxation, anti-necrosis, anti-aging, and anti-apoptotic effects (Nepal *et al.*, 2019). Gastrodin could effectively reduce the level of lipid peroxidation, remove oxygen-free radicals, play an antioxidant role, reduce coupled oxidative phosphorylation, and increase the levels of malondialdehyde and superoxide deaminase (Li *et al.*, 2019). In addition, gastrodin could improve cell

pyroptosis caused by microvascular reperfusion injury via mediating the NLRP3 pathway (de Oliveira *et al.*, 2019). However, the possible effects of gastrodin on TBI, an inflammation-related disease, were still unclear.

The Nrf2 signaling pathway plays a key role in neuroprotection. Astrocytes and neurons of primary cultured Nrf2<sup>-/-</sup> mice were more susceptible to oxidative damage, calcium disorder, and mitochondrial toxicity than wild-type cells (de Oliveira *et al.*, 2019; Hwang *et al.*, 2019). Hwang *et al.* (2019) found that nerve cells pretreated with coffee white fat could activate Nrf2 and its downstream HO-1, and protect nerve cells from Parkinson's related neurotoxin 6-hydroxy dopamine (6-OHDA). It is therefore hypothesized that Nrf2 pathway has potential effects on the inflammation in several diseases. Targeting this signal pathway is important for the treatment of related neurological diseases.

In this study, the effects of gastrodin on a rat model of TBI were investigated. The investigations showed that gastrodin inhibited neuronal apoptosis and suppressed oxidative stress in brain tissues in TBI-induced rats. Gastrodin could serve as a promising and potential drug for the treatment of TBI.

## Materials and Methods

### Traumatic brain injury (TBI) rat model

Sprague–Dawley (SD) rats (Jackson Laboratories, ME, USA) were subjected to TBI model construction. Rats were fed ad libitum and kept under a 12-h/12-h light–dark cycle. All the animal experiments were performed with the approval of the Institutional Animal Care and Use Committee of Qingyang People's Hospital (Gansu, China) (Approval no. 2019141). Chloral hydrate (5%, 0.1 mL/10 g) was used for anesthesia. Rats were fixed to the stereotaxic apparatus, and the skull was widely exposed. TBI model was constructed with 450-g steel weight dropping from 1.5 cm height. Thus, the left cortex was moderately injured. The animals in the sham group were subjected to the same operation except for the dropping of the steel weight. Gastrodin (purity >99.2%) was purchased from Kunming Pharmaceutical Corp (Kunming, Yunnan, China). In TBI+ gastrodin group, after the operation, rats were administrated with the indicated concentration of gastrodin for a week. Thereafter, rats were sacrificed by breaking the neck for further study. Rats in the sham group underwent the same surgery.

### Neurobehavioral evaluation

The neurological score was evaluated with a well-established, neurological severity scoring (NSS), open-field

and Rotarod testing after TBI. The evaluation was carried out without knowing the different treatments. Behavioral tests were repeatedly performed to further validate the outcomes. The NSS test reflects the motor (muscle status, abnormal movement), sensory (visual, tactile, and proprioceptive), balance, and reflex functions of rats. Neurological score ranging from 0 to 18 (0 = normal function; 18 = maximal deficit) represents the neurological lesion.

### Measurement of brain water content

The brain in rats was removed immediately after sacrifice. Then, the brain weight was recorded and then dried in an oven until the weight was unchanged. The dried brain was weighed to get dry weight. The percentage of brain water was calculated as [(Wet weight–Dry weight)/Wet weight] \*100%.

### TUNEL staining

Sliced sections were digested with 20 mg/mL proteinase K at 37°C for 15 min. Then, sections were rinsed in phosphate-buffered saline (PBS) and added with 0.3% H<sub>2</sub>O<sub>2</sub> for 10 min, followed by incubation with 0.1% sodium citrate and 0.1% Triton X-100 solution for 2 min. TUNEL reaction mixture with terminal deoxynucleotidyltransferase (TdT) (Sigma-Aldrich, St. Louis, MO, USA) was added into the sections at 37°C under humidified conditions followed by 4',6-diamidino-2-phenylindole (DAPI) staining. Each image was captured under a confocal microscope.

### Assessment of antioxidant activity

The levels of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GSH-Px), and malondialdehyde (MDA) were spectrophotometrically determined with the commercial kits from Nanjing Jiancheng Bio-engineering Institute (Jiangsu, China) in accordance with manufacturer's instructions.

### Western blot analysis

Brain tissues were homogenized with RIPA lysis buffer (Beyotime Institute of Biotechnology, Jiangsu, China). Supernatants were obtained by centrifugation of homogenates at 12,000 × g at 4°C for 10 min. After the protein concentration was measured by BCA Protein Assay Kit, protein samples were electrophoresed and transferred onto PVDF membranes (EMD Millipore, Billerica, MA, USA). Then, the membranes were blocked with 5%

bovine serum albumin for 1 h, and incubated with the primary antibodies: GAPDH (1:10000; ab8245, Abcam, Cambridge, UK), BCL-2 (1: 1,000; ab32124, Abcam), Bax (1: 1,000; ab32503, Abcam), Cleaved-caspase 3 (1: 1,000; ab32042, Abcam), Nrf2 (1:1000; ab62352, Abcam), H3 (1:1000; ab1791, Abcam), HO-1 (1:1000; ab52947, Abcam), and NQO-1 (1:1000; ab80588, Abcam) for 2 h at room temperature. Subsequently, the membranes were incubated with anti-rabbit or anti-mouse IgG, HRP-linked antibodies at room temperature for 1 h. The blots were analyzed with an ECL kit (Abcam).

### Statistical analysis

Results were displayed as mean  $\pm$  SD. Statistical analysis was performed using GraphPad Prism 5.03 (GraphPad Software Inc., San Diego, CA, USA). Significance was assessed by Student's t-test.  $p < 0.05$  was considered as a significant value.

## Results

### Gastrodin exerts neuroprotective effects on TBI rats

To evaluate the effects of gastrodin on brain deficits in TBI rats, the NSS score in TBI rats after gastrodin treatment was analyzed. As shown in Figure 1(A), compared to rats in the sham group, TBI caused a significantly higher NSS score. The NSS score was reduced in gastrodin-treated TBI rats. Moreover, the water content in the brain was also assessed. TBI stimulation induced elevated brain water content compared to rats in the sham group. However, gastrodin reduced water content in TBI rats (Figure 1B). The results indicated that gastrodin promoted neural injury recovery after TBI establishment.

### Gastrodin inhibits the apoptosis of nerve cells in TBI rats

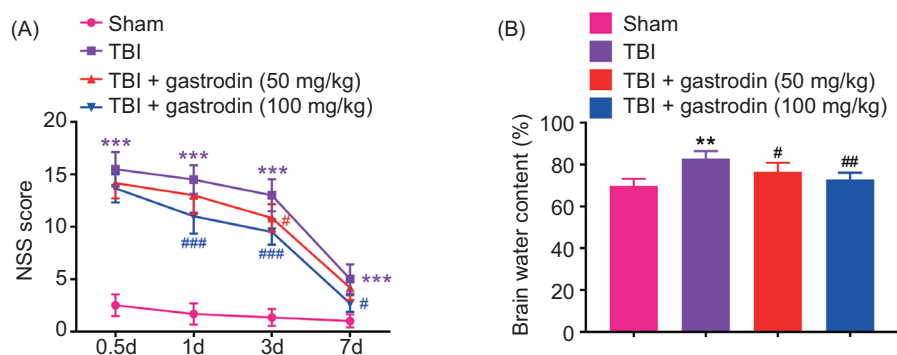
The number of TUNEL-positive cells was calculated to evaluate the effects of gastrodin on the apoptosis of nerve cells. The TUNEL-positive cell number was elevated dramatically in the TBI group. However, significantly fewer TUNEL-positive cells were observed in gastrodin-treated TBI rats (Figure 2A). Additionally, the expression levels of the pro-apoptotic proteins Bax and BCL-2 were measured. TBI induced the protein expressions of Bax and cleaved caspase-3, and suppressed BCL-2 protein expression. However, gastrodin treatment markedly reversed these protein alterations (Figure 3B), suggesting its protective role against nerve cell apoptosis.

### Gastrodin alleviates TBI-induced oxidative stress

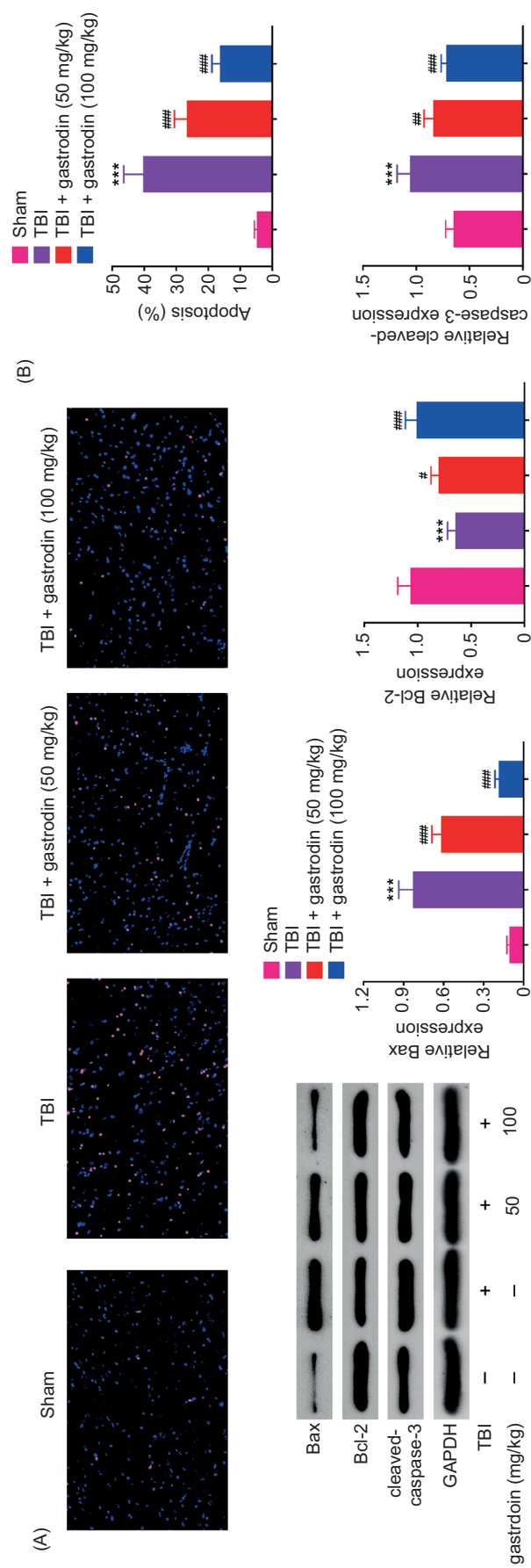
To examine the effects of gastrodin on oxidative stress, the levels of MDA, GSH-px, SOD, and CAT were determined. The levels of SOD, GSH-px, and CAT in the TBI-induced group were significantly decreased, whereas that of MDA was markedly enhanced (Figure 3). Gastrodin greatly exerted the antioxidant capacity, evidenced by increasing the levels of SOD, GSH-px, and CAT, and decreasing the MDA level (Figure 3). These data suggested that gastrodin exerted antioxidative properties in TBI-treated rats.

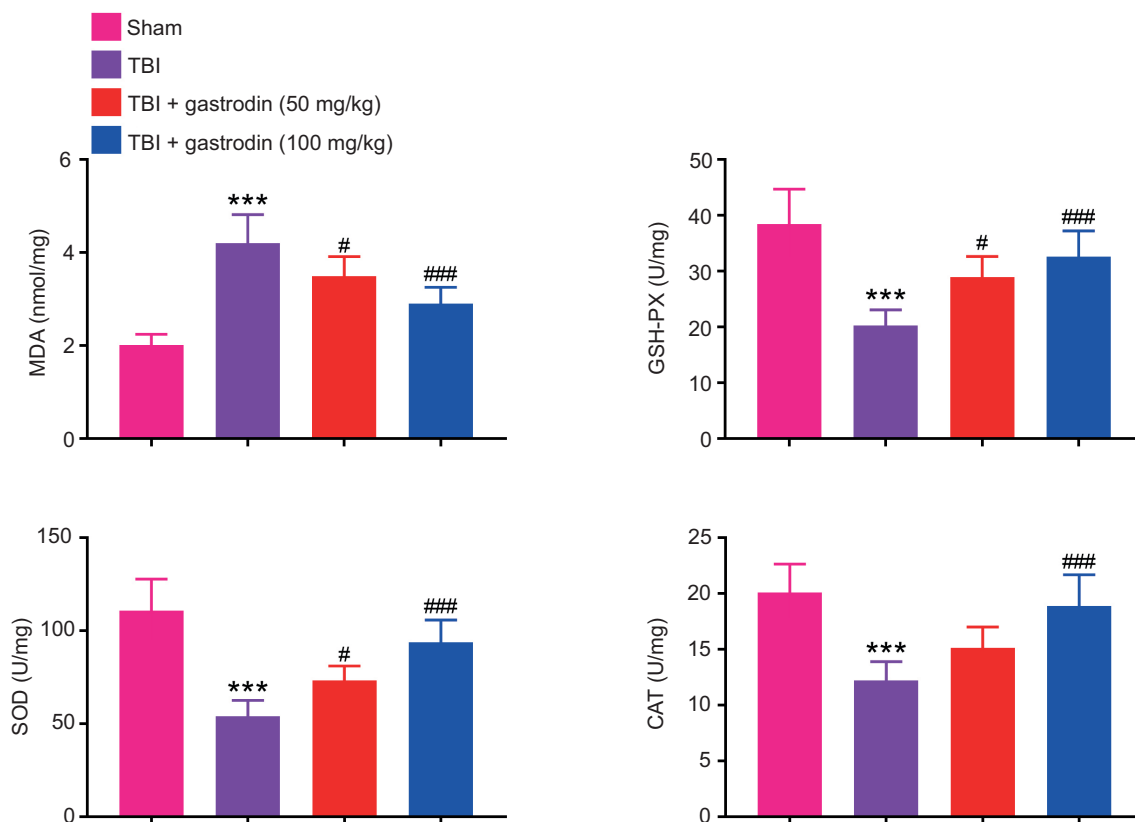
### Gastrodin exerts neuroprotective effects in TBI rats via Nrf2 signaling pathway

To reveal the potential mechanism of gastrodin-mediated effect in TBI rats, the regulatory role of gastrodin in Nrf2 signaling pathway was determined. The Nrf2 signaling pathway was activated in TBI rats, as demonstrated



**Figure 1.** Gastrodin alleviates brain injury in TBI rats. (A) The NSS score of brain in the sham, TBI, TBI + gastrodin 50 and 100 mg/kg groups. (B) The brain water content in the sham, TBI, TBI + gastrodin 50 and 100 mg/kg groups. \*\* $p < 0.01$ , \*\*\* $p < 0.001$  versus sham group; # $p < 0.05$ , ## $p < 0.01$ , ### $p < 0.001$  versus TBI group.





**Figure 3. Gastrodin repressed oxidative stress in the brain of TBI rats. The levels of MDA, GSH-px, SOD, and CAT in the brain of sham, TBI, TBI + gastrodin 50 and 100 mg/kg groups. \*\*\* $p < 0.001$  versus sham group; # $p < 0.05$ , ### $p < 0.001$  versus TBI group.**

by increased levels of Nrf2, HO-1, and NQO-1. The increased levels of Nrf2, HO-1, and NQO-1 were further aggravated by gastrodin treatment (Figure 4). These data suggested that gastrodin exerts neuroprotective effects on TBI rats via Nrf2 signaling pathway.

## Discussion

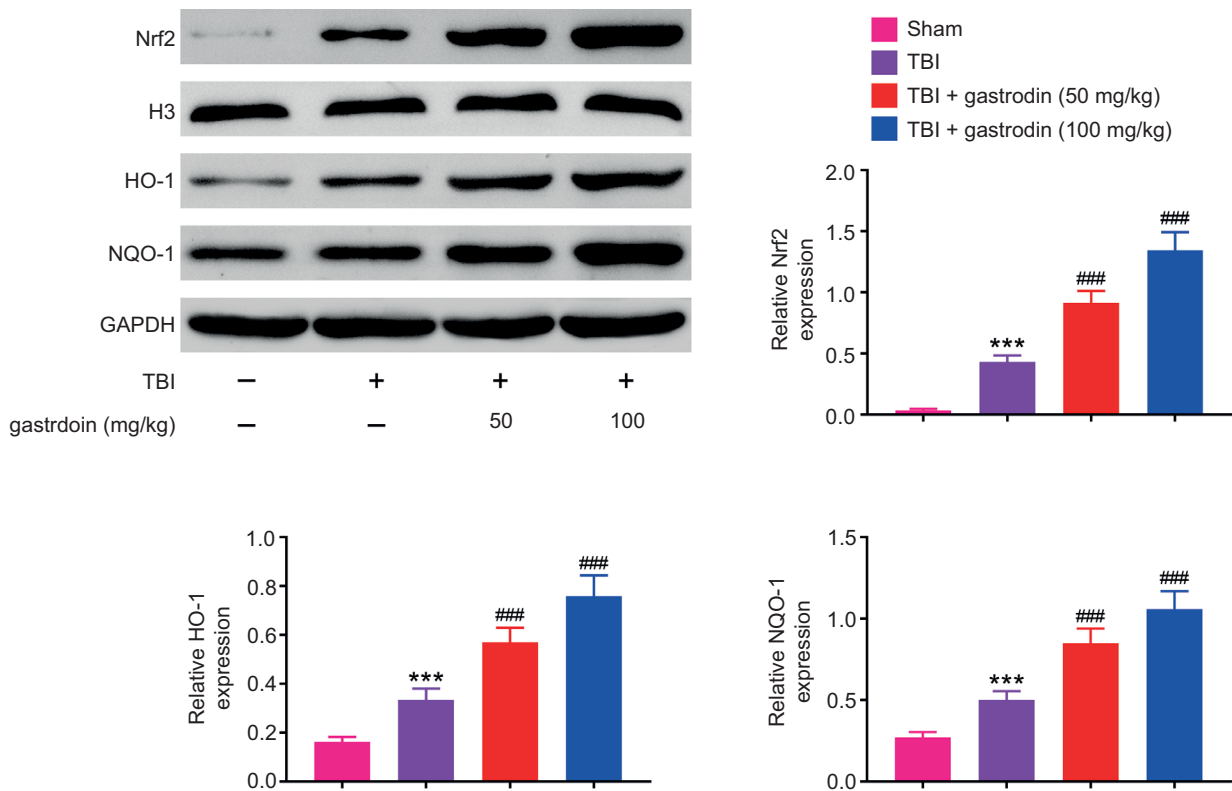
The incidence of TBI has been increasing in recent years. Despite the great progress in research on TBI and improvements in diagnostic techniques and treatment measures, its mortality and disability rates have not decreased significantly (Wearne *et al.*, 2021). Due to its complex diagnosis and treatment, high mortality and disability rates, its mechanism of injury has been one of the hot spots in neurosurgical research (Sowter *et al.*, 2021). A large number of studies have confirmed that brain dysfunction after TBI is not only caused by the primary injury such as the initial mechanical force, but also largely related to the complex “secondary strike” of neurons, namely the secondary neuron injury (Merritt *et al.*, 2021). In this study, gastrodin inhibited neuronal apoptosis and inhibited oxidative stress in brain tissues in TBI rats. The present data provided a new sight into the

pathology of TBI and provided a promising drug for TBI treatment.

Oxidative stress refers to the process of excessive production of highly active molecules such as ROS in the body when the body is subjected to various harmful stimuli, and the imbalance between oxidation system and antioxidant system results in tissue damage (Zhang *et al.*, 2019). After TBI occurs, a large number of free radicals are generated, and oxidative stress plays an important role in the pathophysiological process after TBI. Oxidative stress causes direct damage to nerve cells through lipid peroxidation and protein oxidation. In addition, it can also mediate mitochondria and other pathways to indirectly induce neuronal apoptosis (Yuan *et al.*, 2019). Interestingly, in this study, gastrodin inhibited TBI-induced neuronal apoptosis in rats. In addition, gastrodin inhibited TBI-induced oxidative stress in brain tissues in rats. These data confirmed that this drug had a protective effect against neuronal apoptosis and oxidative stress, two important causes of TBI, proving its potential in treating TBI.

The various biological activities of gastrodin have been widely reported in previous studies. Gastrodin played an important role in maintaining normal neuron





**Figure 4.** Gastrodin suppresses NLRP3 inflammasome signaling pathway in the brain of TBI rats. The protein expression levels of Nrf2, H3, HO-1, and NQO-1 in the brain of sham, TBI, TBI + gastrodin 50 and 100 mg/kg groups. \*\*\* $p < 0.001$  versus sham group; ### $p < 0.001$  versus TBI group.

function (Yang *et al.*, 2020). For example, gastrodin promotes hippocampal neurogenesis in mice following cerebral ischemia via PDE9 pathway (Xiao *et al.*, 2021). Gastrodin could attenuate inflammatory response and cell migration via Notch-1 pathway in activated microglia (Yao *et al.*, 2021). In addition, in the nervous system, gastrodin alleviated the neurocognitive dysfunction of aged mice via suppressing neuroinflammation and promoted the myelination of the central nervous system (CNS) (Lv *et al.*, 2020). Notably, a previous study indicated the nephroprotective effects of gastrodin against lead-induced oxidative stress and inflammation in mice, which was consistent with our findings. Gastrodin also has other biological functions. It could alleviate cerebral ischemia/reperfusion injury via inhibiting pyroptosis. Whether gastrodin could exert protective effects on TBI through its other abilities need further investigation.

In this study, it is noticed that gastrodin upregulated the expression of Nrf2 downstream proteins, suggesting the activation of Nrf2 pathway in brain tissues of TBI rats. The Nrf2 pathway plays a key role in neuroprotection (Chen *et al.*, 2021). It has been reported that several drugs, including gypenosides, provide neuroprotection

effects via Nrf2 pathway (Tian *et al.*, 2021). Astrocytes and neurons of primary cultured Nrf2<sup>-/-</sup> mice were more susceptible to oxidative damage (Dong *et al.*, 2021). These studies, together with our findings, confirmed that the Nrf2 pathway could be a promising target to improve the neuroprotection effects.

In conclusion, the present study found that gastrodin had a neuroprotective effect on rats with TBI by inhibiting neuronal apoptosis and suppressing oxidative stress. Mechanically, gastrodin activated the Nrf2 pathway in brain tissues from TBI rats. Therefore, gastrodin could serve as a promising drug for TBI treatment.

#### Availability of data and materials

All data generated or analyzed during this study are included in this published article.

#### Competing Interests

The authors state that there are no conflicts of interest to disclose.

## Ethics Approval

Ethical approval was obtained from the Ethics Committee of Qingyang People's Hospital (Approval no. 2019141).

## Contribution of Authors

Both Dan Wang and Xiaoqing Dong designed the study and supervised the data collection. Dan Wang analyzed and interpreted the data, Xiaoqing Dong prepared the manuscript for publication and reviewed the draft manuscript. All authors have read and approved the manuscript.

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